

# medilines

## Mucicarmine Colour

### DESIGN TO DEMONSTRATE

Mucin

### REAGENTS & PREPARATIONS

**Mucicar Solution A**

**Mucicar Solution B**

**Mucicar Solution 1:**

Mix A and B in equal volume  
(Solution will remain stable for one  
week. Preferably prepare fresh)

**Mucicar Decolouriser:**

Ready to use

**Mucicar Solution 3:**

Ready to use

**Mucicar Solution 4:**

Ready to use

### TISSUE SAMPLE

4-5µ paraffin sections of 10% neutral buffered formalin fixed tissue.

### STAINING PROCEDURES

1. Remove Paraffin and bring sections to distilled water.
2. Place into the **Mucicar Solution 1** for 1 minute to stain Nuclei.
3. Rinse in distilled water.
4. Decolourise with **Mucicar Decolouriser** for 10 seconds.
5. Wash in running tap water for 1 minute and rinse in distilled water.
6. Place in **Mucicar Solution 3** for 30-40 minutes or longer up to 60 minutes (if a more intense stain is desired) or microwave HI power, 45 seconds
7. Rinse quickly in distilled water.
8. Place in **Mucicar Solution 4** for 10-20 seconds.
9. Rinse quickly in distilled water.
10. Dehydrate in graded alcohols.
11. Clear in xylene, three or four changes.
12. Mount with synthetic resin.

### EXPECTED RESULTS

Mucin:	Deep Rose to red
Capsule of Cryptococcus:	Deep Rose to red
Nuclei:	Black
Other tissue elements:	Yellow or Orange

### NOTES

Allow the Mucicarmine Colour reagent pack to warm to room temperature prior to use.

### Reference

Culling, C.F.A.: *Handbook of Histopathological Technique*, 3rd ed., Butterworths, London, 1974, pp. 304-305.

Luna, L.G.: *Manual of Histologic Staining Methods of the Armed Forces Institute of Pathology*, 3rd ed., New York, McGraw-Hill, 1968, pp. 161-162.

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